



Organic dust exposure induced pulmonary damage among livestock workers.

Dr. Anu Nag¹, Dr. R.S. Sethi², Dr. Akashdeep Singh³

¹Dept. of Human Genetics, Punjabi university Patiala, Punjab

Naganu111@yahoo.co.in

²B.V.Sc & A.H., M.V.Sc, Ph.D., FIAVA, Professor and Head, Department of Animal Biotechnology, College of Animal Biotechnology, Guru Angad Dev Vety & Animal Sciences University, Ludhiana 141004, Punjab, India

Adjunct Professor, Department of Biomedical Sciences University of Saskatchewan, Canada

email: sethi116@gmail.com; rs.sethi@yahoo.com

³Professor & Head, Department of Pulmonary Medicine, DMC&H, Ludhiana.

Received: 19 Apr 2022; Received in revised form: 13 May 2022; Accepted: 22 May 2022; Available online: 29 May 2022

©2022 The Author(s). Published by Infogain Publication. This is an open access article under the CC BY license

(<https://creativecommons.org/licenses/by/4.0/>).

Abstract— Livestock dust contains immunologically potent substances including allergens, endotoxins, microbial compounds, bacteria, fungi, viruses, pathogenic infectious organisms, particulate matter (PM), various poisonous gases such as ammonia, hydrogen sulphide (H₂S), methyl acetate, propanoic acid, heptane etc. It stimulates the immune system through inflammatory and allergenic microbial agents (molds, bacteria, virus and allergens) and microbial-associated molecular patterns (e.g., endotoxin, glucans and peptidoglycans), to result in inflammatory reactions. Farmers are at the risk of developing airway diseases resulting from repeatedly exposures on the livestock farms. There is a paucity of data on in vivo and in vitro cellular and molecular changes following multiple exposures to these livestock contaminants and their long-term impact on the environment as well as human health. The mechanisms of lung dysfunction are still largely unknown. So, there is strong need to look at the combined effect of all the components of livestock dust as stimulatory factors for respiratory hazards. The development of preventive strategies to reduce exposure will be required- in-depth and identification of factors that affect day-to-day variability in exposure.

Keywords— Organic dust, livestock dust, endotoxins, respiratory disorders, pulmonary health.

INTRODUCTION

Livestock keeping is an age-old practice in India and it is the second income generating activity after crop in the agricultural economy of India. Livestock production and agriculture are intrinsically linked to each other and both are crucial for overall food security. About 20.5 million people depend upon livestock for their livelihood. This sector provides employment to about 8.8 % of the population in India and contributes 4.11% GDP of the total Agriculture GDP (25.6%). Unfortunately, livestock farmers are at the risk of developing various diseases associated with exposures to livestock dust. Livestock farming is one of the agricultural industries where farmers

are at increased risk for respiratory health problems (Merchant et al. 1991).

Organic dust availability and its composition

In the field of occupational hygiene, airborne and settled particulate material of biologic origin is often referred as organic dust. Organic dusts contain particles of plant, animal and microbial origin (Douwes et al., 2003). Dust exists with heterogeneous composition and this is predominantly organic from animal confinement (Donham, 1986). Organic dusts found in agricultural environments display equally complex composition, although the primary sources may differ. Rylander (1994) reported in animal confinement buildings some organic dust particles which originated from the

animal feed, but the main sources of microorganisms, allergens, and toxins are animal dander, urine, and feces. Microorganisms are widespread in the environment and are often a major component of organic dusts because of the nutrients that the dusts contain. The microflora of organic dust depends on the source material, which depends in turn on a variety of factors, among them substrate composition, acidity, aeration, water availability, and temperature etc. (Lacey, 1994). The main agent in organic dust is endotoxin, a major component of the outer membrane of Gram-negative bacteria, but components of Gram-positive bacteria (peptidoglycans) and fungi (glucans) are also present. The organic fraction of these dusts may contain yeasts, molds, mesophilic and thermophilic bacteria (G-positive and G-negative), histamine, cow urine antigen, mite antigen, endotoxins and pharmaceutical compounds (Donham, 1986; Kullman et al. 1998; Kemper, 2008). Similarly, an “immunoallergic” and inflammatory reaction following the inhalation of organic dust contained in hay among dairy workers due to endotoxins has been proposed to explain this phenomenon (Dalphin et al., 1993; Eduard et al., 2009; Poole and Romberger, 2012). A large number of studies suggested that total dust is comprised of variety of factors such as , insect parts etc. which means the dust is biologically active and will react to the defense system of the respiratory organs.

particle size, concentration of bacterial products LPS and PGN etc. and revealed relevant assessment techniques for the measurement of mold spores, mycotoxins, bacteria, allergenic proteins, endotoxins, and microorganisms in the organic dust (Kiryckiet al. 2010 and Hawley et al. 2015).

Noxious Gases also act as potential health hazards along with these dust particles such as ammonia which is present inside unbedded dairy calf and hydrogen sulphide which is very poisonous is produced inside the farms during manure agitation (Eduard et al. 2017). Heedreiket al. 2007 and Guidry et al. 2018 also demonstrated that in animal production farms, the dusts from the animals, their feed, and their faeces contains high levels of poisonous gases such as ammonia (NH₃) which comes primarily from the animal’s urine and faeces, and hydrogen sulphide (H₂S) from manure pits, especially during agitation and emptying. This fact is well supported by Eduard et al. 2017 and May et al. 2012 that livestock buildings may contain concentrations of contaminants that can negatively affect human health.

As mentioned, in table no. 1. The dust in livestock barns is comprised of molds, actinomycetes, dried fecal matter, pollens

Table 1. Classification of dust inside the livestock barns:

Source of dust	Type of dust	Cause of dust
Grain	Molds, actinomycetes	Storage problem
Hay	Molds, actinomycetes	Poor conservation
Straw	Molds, actinomycetes	Combining/poor conservation
Silage	Molds	Poor conservation
Animal Debris	Faeces, urine, hair, skin, feathers Fungi, bacteria	Animal activity, barn, cleaniless, ventilation, etc.
Feeds	Numerous particles	Feed-distribution/poor ventilation

Source: Air Quality Inside Livestock Barns (gov.on.ca)

Epidemiological Studies

Respiratory disease is an important clinical problem for livestock farmers which significantly increased risk of their respiratory morbidity and mortality. Farmers and other individuals involved in livestock farming have an increased risk for acute and chronic respiratory disorders; their respiration is routinely challenged by intense exposure to several chemical and biological substances such as odorous gases, and organic and inorganic dusts (Schenker et al. 1998; Kirkhorn et al. 2000; Omland; 2002; Sigsgaard et al. 2010). The dairy farmers are exposed to high concentrations of organic dusts, this increased risk

seems mainly driven by chronic exposure to these dusts (Basinas et al., 2014; Pfister et al., 2018).

Various studies involving large numbers of exposure measurements which have been published previously for dairy and swine confinement industries have shown that organic dust exposures may vary qualitatively as well as quantitatively from one occupation to another but these dusts had stronger respiratory effects on the lungs of the farmers (Huyet al. 1991, Smidet al. 1992; Preller et al. 1995; Kullman 1998). Kouimintziset al. 2007 and Szczyreket al. 2011 found a strong association between developing bronchitis, COPD and lung reduced force

expiratory volume among livestock farmers who are exposed to high levels of organic dust in confined buildings. Dalphin *et al.* 1989 studied the prevalence of bronchitis and reduced respiratory function among livestock dairy farmers. He further concluded that dairy farmers are at a greater risk of developing respiratory

disfunction. A large number of studies also supported and found a strong potential association between animal farming and non-malignant disorders, like asthma, chronic bronchitis organic dust toxic syndrome and hypersensitivity pneumonitis (Zejda *et al.* 1993; Omland 2002; Hoppin *et al.* 2003) as shown in table 2.

Table 2. Respiratory conditions due to occupational exposure

OCCUPATION	EXPOSURE	RESPIRATORY CONDITIONS
Dairy farmers	Organic dust Feed additives Thermophilic bacteria and fungi Fungi Microbial toxins Storage mites Irritant gases	Chronic Bronchitis, Organic dust toxic syndrome Asthma Allergic alveolitis (farmer's lung) Bronchiolitis Toxic pneumonitis (silo filler's lung) Rhinitis COPD (chronic obstructive pulmonary disease)

Source: Merchant JA et al. Textbook on Clinical Occupational and Environmental Medicine. Eds. Rosenstock L, Cullen, 1994.

Various studies demonstrated that organic dusts present particularly in Concentrated Animal Feeding Operations (CAFOs) i.e., is a system of farming which concentrates a large number of animals into a small space for maximum efficiency, may cause lung inflammation and assessed the respiratory health of the workers by testing dust mitigation

strategies and further reported that organic dust and its components may be a deciding factor in understanding inflammatory response among livestock farmers who are exposed to the environment (Senthilselvan *et al.* 1997; Schneberger *et al.* 2016 and Hawley *et al.* 2015) as shown in fig. 1.

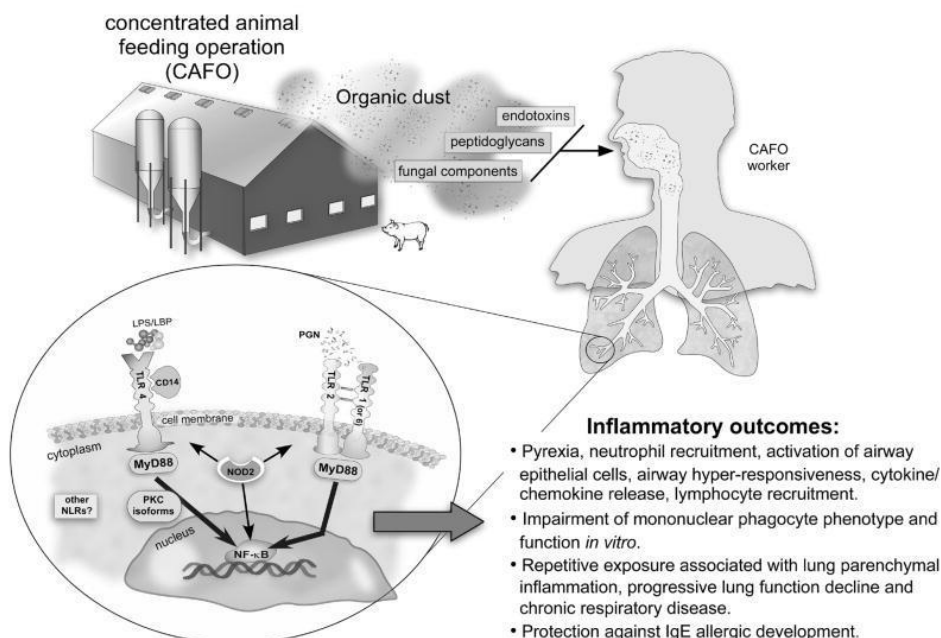


Fig.1: Poole, J. A., & Romberger, D. J. (2012). Immunological and inflammatory responses to organic dust in agriculture. *Current opinion in allergy and clinical immunology*. Diagrammatic presentation of the agents and immunologic and inflammatory consequences of organic dust exposure in the agriculture industry in large, concentrated animal feeding operations (CAFOs).

Exposure studies found a strong association between COPD, dust and endotoxins in confined livestock buildings and revealed the fact that livestock farmers had significant higher mortality rates due to their respiratory illness (Monsoet *et al.* 2004; Szczyrket *et al.* 2011). Van *et al.* 2016 and Sigsgaard *et al.* 2020 also recently investigated the impact of livestock farming on lung health of farmers in their epidemiological studies and reported the risk of developing chronic bronchitis and bronchial obstruction/COPD among livestock farmers. They concluded the higher prevalence of lower respiratory tract infections in the area with a high density of livestock farms. Dairy farm dusts are complex mixtures that contain both toxic and immunogenic compounds. Dust exposure is a major source of respiratory morbidity and mortality among agricultural workers (American Thoracic Society, 1998; Schenker, 2000; Linaker and Smedley, 2002; Cleave *et al.* 2009; Reynolds *et al.* 2013). A large number of studies indicated that dairy workers, in particular, have increased risks for **asthma, rhinitis, sinusitis, mucus membrane inflammation syndrome, bronchitis, chronic obstructive pulmonary disease (COPD), hypersensitivity pneumonitis, and organic dust toxic syndrome** (Kullman *et al.* 1998; Gainet *et al.* 2007; May *et al.* 2012; Reynolds *et al.* 2013).

Taluja, *et al.* 2019 also reported that occupational exposure of dust causes respiratory hazards to farm workers and concluded that dust adversely affects the respiratory function by deteriorating the lung function parameters. Similarly, number of exposure studies reported a range of respiratory and other symptoms including bronchitis, chest tightness, nasal congestion, organic dust toxic syndrome, occupational asthma, mucus membrane irritation, nausea, headache, mood changes, altered immunity among workers who are exposed to organic dust (Sahlander *et al.* 2012; Viegas *et al.* 2013; Iowa Concentrated Animal Feeding Operations Air Quality Study, 2018) and increased risks of lung cancer (Peters *et al.* 2012).

Various studies indicated health hazards and recognized a number of syndromes among workers in intensive livestock production (Donham *et al.* 1995; Hartung and Schulz, 2008). Organic dust toxic syndrome recognized in workers is predominantly a severe flu-like syndrome originally described in farmers and other persons who are occupationally exposed to dusty conditions. (Donham *et al.* 2002; Hartung and Schulz, 2008) again reported the prevalence of ODTS ranges from 10 to 30% in workers, depending on the type of intensive animal production and use of facilities. Several studies have indicated that the prevalence of persistent restriction of airflow and pulmonary disorders are higher among dairy farmers than in the general population (Stoleski *et al.*, 2015; Guillien

et al., 2016; Marescaux *et al.*, 2016; Guillien *et al.*, 2019). As dairy farmers are exposed to high concentrations of organic dusts, this increased risk seems mainly driven by chronic exposure to these dusts (Basinas *et al.*, 2014; Pfister *et al.*, 2018). These findings suggest that presence of multiple microbial and non-microbial factors in OD can cause broad-range of health effects upon exposure which is a significant public health concern.

Mechanism involving lung inflammation

Cytokines play an important role in determining pulmonary diseases which includes **platelet-derived growth factor (PDGF), Interleukin-1 (IL-1), transforming growth factor- β (TGF- β), tumor necrosis factor- α (TNF- α), Insulin like growth factor I (IGF-I), Interleukin- γ (IL- γ) 1** (Kelley, 1990). Alterations in cytokine production, secretion, and action represent determining forces in the destructive and inflammatory lung disorders as shown in fig. 2.

Recognition of micro-organisms is an important function of the innate and adaptive immunity and pattern recognition receptors (PRRs) recognize a variety of pathogen-associated molecular patterns (PAMPs) from viruses, bacteria, fungi and parasites (Takeda *et al.* 2003). Toll-like receptors (TLRs) comprise a group of PRRs which are expressed by most immune cells. When binding to PAMPs, immune responses including expression of proinflammatory cytokines and type I interferons (IFNs) are mediated through activation of the transcription factor **NF- κ B and interferon regulatory factors (IRFs)**. Currently, eleven TLRs have been identified in humans of which TLR2 and TLR4 are the most studied. Toll-like receptor 4 is activated by the endogenous molecules or danger signals released during tissue injuries in addition to bacterial endotoxins (Akira *et al.* 2006; Tsan and Gao 2004). Thus, TLR4 is the prototypical sensor of infection or injury that arranges the innate response via a sequential activation of both cell surface and endocytic signaling pathways (Ghosh *et al.* 2015). The increased expression of TLR4 has been associated with the lung dysfunction and elevated TLR4 subsequently activates nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) which in turn directs the expression of various chemokines and pro-inflammatory cytokines including Interleukins-1 β and tumor necrosis factor TNF- α (Strieter *et al.* 2002).

Lipopolysaccharides (LPS) is a major component of the outer membrane of gram-negative bacteria, (Sugiyama *et al.* 2008) is associated with the development and progression of various lung diseases characterized by chronic inflammatory conditions (Vernooy *et al.* 2002). Single LPS exposure is sufficient to rapidly recruit

neutrophils to the lung and to produce pro-inflammatory cytokines and chemokines (Deetz *et al.* 1997). The recognition of LPS is modulated by soluble factors, such as LPS binding protein and surfactant proteins that are present in airway lining fluid and influence the presentation of LPS to membrane-bound CD14 (Skerrett *et al.* 2004). Binding of LPS to CD14 triggers intracellular signaling that is mediated by Toll-like receptor 4 (TLR4) in association with a secreted cofactor, MD-2 (Akira *et al.* 2003). TLR2 recognizes PAMPs from Gram-positive bacteria like peptidoglycans (PGN) and lipoproteins and binds LPS from special bacteria strains and PAMPs from fungi (Takeda *et al.* 2003 and Akira *et al.* 2006). Zahringert *et al.* 2008 reported that TLR 2 is typically associated with the detection of peptidoglycans,

lipoteichoic-acid, lipoproteins, lipopeptides and zymosan. (Oliveira-Nascimento *et al.* 2012; Kang *et al.* 2009; Oosting *et al.* 2014) further demonstrated the dimerization property of TLR2 with TLR1, TLR6 or TLR 10 which makes it to detect increased range of possible ligands. These studies have shown that TLR2 may be even more important than TLR4 in determining the response to these organic dusts. (Martin *et al.* 1996) also supported in their study that TLR2 may be more important than TLR4 in determining the response during organic dusts exposure. Whereas other studies supported the role of protein kinase C (PKC), TLR9 (Schneberger *et al.* 2016); TLR2 (Poole *et al.* 2011); MyD88 (Bauer *et al.* 2013) in several innate signaling pathways in response to organic dust exposure.

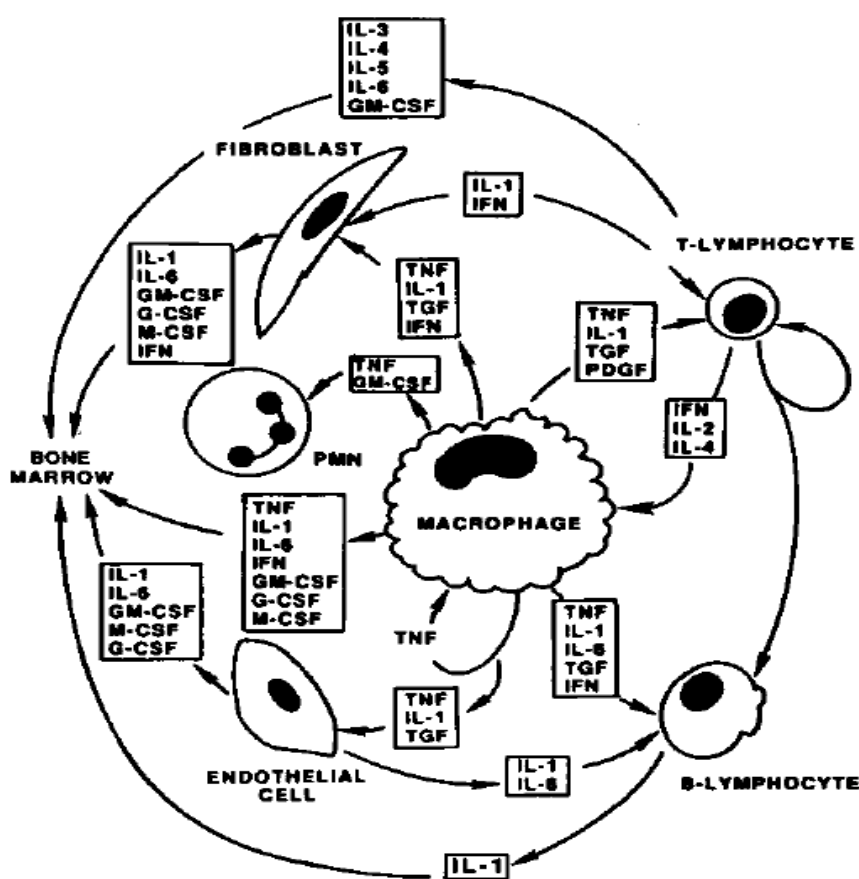


Figure 2. Kelley, J. (1990). Cytokines of the lung. *Am Rev Respir Dis*, 141(3), 765-788.

Schematic showing multiple potential cellular interactions involving cytokines.

Interleukin-1 β is a potent pro-inflammatory cytokine that is crucial for host-defense response to infection and injury (Lopez *et al.* 2011). Interleukin-1 β , encoded by the IL-1 β gene, has been associated with chronic inflammation and plays an important role in lung inflammatory diseases including lung cancer. Elevated levels of IL-1 proteins, in

particular IL-1 β greatly enhance the intensity of the inflammatory response (Bhat *et al.* 2014). (Zahringert *et al.* 2008) reported that TLR2 is associated with the detection of peptidoglycans, lipoteichoic acid, lipoproteins and zymosan. Senthilselvan *et al.* 2009 revealed the association of TLR4 gene and its ligand LPS as an important factor in

lung dysfunction after barn exposure in humans. (Sahlander *et al.* 2010) studied association between T-helper (th) cell, cytokine *profile* and acute response to pro-inflammatory stimuli as markers. Whereas (Kang *et al.* 2009 and Oosting *et al.* 2014) reported that TLR2 can dimerize with TLR1, TLR6, TLR10 and can detect various range of possible ligands. Romberger *et al.* 2015 discovered **proteolytic properties that can stimulate protease activated receptors (PARs) 1 and 2** in the bronchial epithelium and focused on the contribution of proteases to lung inflammation and pathways which contribute to the lung inflammatory response to dust exposure. These proteolytic properties of organic dusts require further investigations because some chemokines are also produced on cleavage although study suggested main effect is through PAR receptors.

Surfactant proteins (SP) are also important constituents of pulmonary defense and are a lipoprotein complex of phospholipids and apoproteins which is a complex critical component in lung immune host defense (Pastva *et al.* 2007). Surfactant, a lipoprotein. Pulmonary SPs are also involved in the pathophysiology of lung injury following exposure to pesticides (Gil *et al.* 2007). Although surfactants appear not to have been studied extensively in terms of organic dust whereas Scheneberger *et al.* 2018 examined the role of the immune surfactant or collectin surfactant protein D (SP-D) in lungs induced prolonged exposure to organic barn dusts and concluded that organic barn dust can reduce lung SP-D, thus leaving workers potentially at risk for a host of pathogens. Sethi *et al.* 2017 also found SP-A and SP-D reductions in A549 lung cell lines exposed to hog barn dusts. This further suggests that some of the inflammatory response generated in lungs to these dusts might be attributable to reductions in these immune surfactants.

In- vivo studies

Multiple researches identified organic dusts as a major problem in lung inflammation and have shown that organic dust exposure on human subjects and animals are helpful in accessing the pulmonary health of the occupational workers (Senthilselvan *et al.* 1997; Charavarymath *et al.* 2005; Scheneberger *et al.* 2016). Various animal models have been designed by the researchers which helped in understanding the molecular and cellular mechanisms of lung inflammation following exposures to livestock dust (Sethi *et al.* 2013 and Charavaryamath and Singh 2006).

Charavaryamath *et al.* 2005 designed first rat model of human exposure to barn air which was mimicked human exposure of fulltime barn workers (8h/day, Monday to Friday) and found that single and multiple exposures to endotoxin rich-swine barn air induce lung inflammation

characterized by infiltration of inflammatory cells, increased mucus positive-epithelial cells in addition to the presence of endotoxins, dust, ammonia, microorganisms, aeroallergens.

Charavarymath *et al.* 2006 supported that PIMMS cells are important because they have shown to produce both pro and anti-inflammatory cytokines to regulate lung inflammation. Further Gamage *et al.* 2007 demonstrated the role of pulmonary intravascular monocytes/macrophages (PIMMS) recruitment and their functions in lung inflammation induced following exposure to the barn air by using a rat model. Further interactions between PAR2 and TLR4 were also well demonstrated by Rallabandiet *et al.* 2008. Lundin and Checkoway, 2009 reported altered TLR expression among livestock farmers who were probably exposed to organic agents and concluded that reduced TLR expression may be related to the increased prevalence of respiratory disorders among livestock farmers.

Sahlander *et al.* (2010) hypothesized that T-helper (Th) cell cytokine profile and acute response to pro-inflammatory stimuli as immune response among pig farmers and smokers. After in vivo exposure, altered TLR expression was only observed in controls and the ex vivo stimulations showed an attenuated response in farmers compared to the control group. The inflammatory systemic response to pro-inflammatory stimuli is altered in farmers probably because of adaptive mechanisms arising from chronic exposure to organic material and microbial compounds, have an increased number of inflammatory cells in peripheral blood and an increased proportion of circulating Th2- type lymphocytes which, among pig farmers, were positively related with the duration of work in pig confinement buildings.

Sethi *et al.* 2013 revealed a decrease in Clara cells and an increase in numbers of mucus producing goblet cells in the airway epithelium in exposed knockout TLR9^{-/-} animals after chronic barn exposure. Scheneberger *et al.* 2016 reported that in TLR9 knockout mice no stronger responses has been detected as compared to TLR2 and TLR4 mice when exposed to the barn environment. Sethi *et al.* 2017 further demonstrated the role of innate inflammatory mechanism, especially TLR4 and TLR9 following exposure to dust and other pollutants in agriculture environments by using in vitro models and laboratory animals.

Nath *et al.* 2018 illustrated the association between the exacerbated inflammation upon persistent barn organic dust exposure, pathogenesis of lung inflammation and lung function decline. He investigated the inflammatory effects of Barn Dust Extracts on human lung epithelial (BEAS2B)

and macrophage (THP-1 monocyte derived) cell lines on a kinome array to determine phosphorylation events in the inflammatory signaling pathways. Further he concluded that workers exposed to persistent barn air are at occupational risk of developing a range of respiratory illnesses and there is a critical need to understand the mechanisms behind those illness.

Kaur and Sethi, 2020 concluded that multiple exposures of poultry barn air resulted in lung damage and damage is more severe when combined with LPS. Multiple exposures in combination with LPS significantly altered the mRNA and protein expression of TLR-4 and IL-1 β . Expression of TLR4 and IL-1 β showed a synergistic effect when combined with LPS. Woldeamanuel *et al.* 2020 and Soumagneet *et al.* 2020 reviewed that dairy farmer could be exposed to various potentially hazardous exposures that can cause respiratory symptoms due to higher inflammatory markers such as such as TNF α , fibrinogen, IL-6 and CRP

In vitro studies

Poole et al. 2008 found that repetitive organic dust exposure significantly decreases markers of antigen presentation and host defense function in monocyte-derived macrophage (MDMs). Bacterial cell components appear to be driving these impaired responses. **Expression of HLA-DR, CD80, and CD86;** phagocytosis; and intracellular bacterial killing were significantly decreased with organic dust exposure (ODE)-challenged versus control MDMs. Responses were retained after marked depletion of endotoxin. PGN, LPS, and PGN plus LPS significantly reduced MDM surface marker expression and, except for LPS alone, also reduced phagocytosis. ODE-challenged MDMs had significantly diminished cytokine responses (TNF- α , IL-6, and IL-10) after repeat challenge with high-dose ODE and concluded that organic dust-induced macrophage dysfunction might be important in respiratory disease development.

Viegaset *et al.* 2017 chose macrophages for the in vitro assessment because they are known to be responsible for first-line protection and also for triggering the inflammatory response via secretion of signaling molecules. The cell viability and the inflammatory response, as measured by the production of pro-inflammatory cytokines tumor necrosis factor- α (TNF α) and interleukin-1 β (IL-1 β), are determined in human macrophages derived from THP-1 monocytic cells. This study emphasizes the importance of measuring the organic dust/mixture effects in occupational settings and suggests that differences in the organic dust content may result in differences in health effects for exposed workers.

Nordgren and Charavaryamath in 2018 demonstrated the application of a kinome array to delineate key inflammatory signaling pathways activated upon swine barn dust extract SBDE exposure in vitro. They concluded that SBDE-mediated pro-inflammatory effects are predominantly due to the induction of neutrophilic chemokine IL-8. Differentially phosphorylated peptides implicated in IL-8 induction in BEAS2B cell line include, **TLR2, 4, 5, 7, 8, 9, PKC, MAP kinases (p38, JNK), inflammasomes (NLRP1, NLRP3), NF- κ B and AP-1** due to persistent barn organic dust exposure which is a key contributor to the pathogenesis of lung inflammation and lung function decline.

Nordgren *et al.* 2013 identified the role of mediators derived from polyunsaturated fatty acids exhibit anti-inflammatory, pre-resolving actions and tested the potential of one of these mediators i.e., **maresin-1 (MaR1)**. As bronchial epithelial cells (BECs) play a pivotal role in initiating organic dust-induced inflammation and they investigated the in vitro effects of MaR1 on a human BEC cell line (BEAS-2B) in reducing organic dust-associated airway inflammation. Again Nordgren *et al.* 2015 supported the previous study that the specialized proresolving lipid mediator **maresin-1 (MaR1)** reduced proinflammatory cytokine release and **intracellular adhesion molecule-1 (ICAM-1)** expression in bronchial epithelial cells exposed to extracts of organic dust (DE) derived from swine confinement facilities in vitro. The data suggested that MaR1 might contribute to an effective strategy to reduce airway inflammatory diseases induced by agricultural-related organic dust environmental exposures.

Occupational lung diseases caused and aggravated by various organic and inorganic inhaled dust, fumes, and mist. Therefore, occupational history of livestock farmers should be considered when evaluating respiratory symptoms. The best way to treat OLDs as early as possible are prevention and early detection by controlling the working environment and conducting regular surveillance of workers (Lee, 2011).

There is an urgent need to improve knowledge on several topics, including 1) understanding the nature and pathophysiology of respiratory diseases, 2) diagnostic approaches, 3) disease behavior and natural history, and 4) therapeutic approaches. Key questions on pathophysiology includes genetic susceptibility and both host and environmental factors. For diagnostic approaches, important needs include the validation and standardization of questionnaires, BAL lymphocytosis threshold, specific antibodies, and biomarkers. In addition, new techniques

like genomic classifiers and artificial intelligence to improve diagnosis and prognosis need to be assessed. For example, the diagnosis of hypersensitivity pneumonitis (HP) and distinguishing it from other forms of interstitial lung diseases. The diagnosis of HP relies on the integration of multiple domains such as clinical assessment of exposure, imaging, bronchoalveolar lavage lymphocytosis and histopathological findings. Among patients with fibrotic HP, the MUC5B (mucin 5B) promoter polymorphism is more prevalent than in the general population and is associated with shortened survival (Ley, 2017). Among sensitized individuals, the immune reaction after exposure to an antigen appears to consist of both humoral (i.e., antigen-specific IgG antibodies) and T-helper cell type 1 (Th1) cellular immune responses (Hisauchi-Kojima, 1999). These responses lead to a predominantly lymphocytic inflammatory pattern and granulomatous inflammation (Vasakova et al. 2019). Neutrophilic inflammation may play a role early in the disease whereas impaired function of T regulatory cells may play a role in the exaggerated immune response (Pardo et al. 2000). Similarly, other pulmonary diseases can be detected by analyzing cytokine production, presence of IgG and IgE antibodies. Many other antigens in organic dusts associated with these diseases have not been adequately characterized and better understanding of the mechanisms by which these materials cause clinical disease is necessary.

CONCLUSION

From this review it has been concluded that pulmonary damage risk is associated with raising livestock animals due to organic dust exposure. Once the lung is damaged by organic dusts, regeneration of lung to normal state is almost impossible. Thus, prediction and early diagnosis of lung diseases are important and imperatively necessary. In spite of advanced technologies, overall level of organic dust exposure of livestock workers is very high which poses a serious health threat. Exposure to organic dust and its health effects among workers have been investigated in numerous epidemiological and exposure-assessment studies during the last three decades. Further, multiple components within the dust such as endotoxins, LPS and peptidoglycans bind to a wide variety of innate immune receptors but we still have no clear definition of the potential determinants of personal exposures in livestock farming environments. Work-related respiratory diseases are major contributors to the global burden of respiratory diseases. This review is focused on the findings which suggested that livestock farmers are exposed to high levels of dust and endotoxin consistent with an increased risk of developing respiratory symptoms and diseases.

REFERENCES

- [1] Akira, S. (2003). Toll-like receptor signaling. *Journal of Biological Chemistry*, 278(40), 38105-38108.
- [2] Akira S, Uematsu S, Takeuchi O (2006). Pathogen recognition and innate immunity. *Cell* 2006; 124: 783–801.
- [3] American Thoracic Society. 1998. Respiratory health hazards in agriculture. *Am. J. Respir. Crit. Care Med.* 11/01: S1–S76. *Barns. Am. Ind. Hyg. Assoc. J.* 59: 403–413.
- [4] Basinas, I., Sigsgaard, T., Erlandsen, M., et al., 2014. Exposure-affecting factors of dairy farmers' exposure to inhalable dust and endotoxin. *Ann. Occup. Hyg.* 58, 707–723. doi: 10.1093/annhyg/meu024.
- [5] Bauer, C., Kielian, T., Wyatt, T. A., Romberger, D. J., West, W. W., Gleason, A. M., & Poole, J. A. (2013). Myeloid Differentiation Factor 88–Dependent Signaling Is Critical for Acute Organic Dust–Induced Airway Inflammation in Mice. *American journal of respiratory cell and molecular biology*, 48(6), 781–789.
- [6] Berahovich, R. D., Miao, Z., Wang, Y., Premack, B., Howard, M. C., & Schall, T. J. (2005). Proteolytic activation of alternative CCR1 ligands in inflammation. *The Journal of Immunology*, 174(11), 7341–7351.
- [7] Bhat IA, Naykoo NA, Qasim I, et al. Association of interleukin 1 beta (IL-1 β) polymorphism with mRNA expression and risk of non-small cell lung cancer. *Meta Gene*. 2014; 2:123–133.
- [8] Charavaryamath, C., & Singh, B. (2006). Pulmonary effects of exposure to pig barn air. *Journal of occupational Medicine and Toxicology*, 1(1), 1–4.
- [9] Charavaryamath, C., Janardhan, K. S., Caldwell, S., & Singh, B. (2006). Pulmonary intravascular monocytes/macrophages in a rat model of sepsis. *The Anatomical Record Part A: Discoveries in Molecular, Cellular, and Evolutionary Biology: An Official Publication of the American Association of Anatomists*, 288(12), 1259–1271.
- [10] Charavaryamath, C., Janardhan, K. S., Townsend, H. G., Willson, P., & Singh, B. (2005). Multiple exposures to swine barn air induce lung inflammation and airway hyper-responsiveness. *Respiratory research*, 6(1), 1–13.
- [11] Cleave, J., Willson, P. J., Town, J., & Gordon, J. R. (2010). Fractionation of swine barn dust and assessment of its impact on the respiratory tract following repeated airway exposure. *Journal of Toxicology and Environmental Health, Part A*, 73(16), 1090–1101.
- [12] Dalphin, J. C., Bildstein, F., Fernet, D., Dubiez, A., & Depierre, A. (1989). Prevalence of chronic bronchitis and respiratory function in a group of dairy farmers in the French Doubs province. *Chest*, 95(6), 1244–1247.
- [13] Dalphin, J. C., Debieuvre, D., Pernet, D., Maheu, M. F., Polio, J. C., Toson, B., ... & Depierre, A. (1993). Prevalence and risk factors for chronic bronchitis and farmer's lung in French dairy farmers. *Occupational and Environmental Medicine*, 50(10), 941–944.
- [14] Deetz, D. C., Jagielo, P. J., Quinn, T. J., Thorne, P. S., Bleuer, S. A., & Schwartz, D. A. (1997). The kinetics of grain dust-induced inflammation of the lower respiratory

- tract. American journal of respiratory and critical care medicine, 155(1), 254-259.
- [15] Donham, K. 1986. Hazardous agents in agricultural dusts and methods of evaluation. *Am. J. Ind. Med.* 10:205–220.
- [16] Donham, K. J. (2000). The concentration of swine production: Effects on swine health, productivity, human health, and the environment. *Veterinary Clinics of North America: Food Animal Practice*, 16(3), 559-597.
- [17] Donham, K. J., Cumro, D., & Reynolds, S. (2002). Synergistic effects of dust and ammonia on the occupational health effects of poultry production workers. *Journal of agromedicine*, 8(2), 57-76.
- [18] Donham, K. J., Reynolds, S. J., Whitten, P., Merchant, J. A., Burmeister, L., & Popenorf, W. J. (1995). Respiratory dysfunction in swine production facility workers: Dose-response relationships of environmental exposures and pulmonary function. *American journal of industrial medicine*, 27(3), 405-418.
- [19] Douwes, J., Thorne, P., Pearce, N., & Heederik, D. (2003). Bioaerosol health effects and exposure assessment: progress and prospects. *The Annals of occupational hygiene*, 47(3), 187-200.
- [20] Eduard, W., Pearce, N., & Douwes, J. (2009). Chronic bronchitis, COPD, and lung function in farmers: the role of biological agents. *Chest*, 136(3), 716-725.
- [21] Gainet M, Thaon I, Westeel V, Chaudemanche H, Venier AG, Dubiez A et al (2007) Twelve-year longitudinal study of respiratory status in dairy farmers. *Eur Respir J* 30(1):97–103
- [22] Gamage, L. N. A., Charavaryamath, C., Swift, T. L., & Singh, B. (2007). Lung inflammation following a single exposure to swine barn air. *Journal of Occupational Medicine and Toxicology*, 2(1), 1-12.
- [23] Gil HW, Oh MH, Woo KM, Lee EY, Oh MH, Hong SY (2007) Relationship between pulmonary surfactant protein and lipid peroxidation in lung injury due to paraquat intoxication in rats. *Korean J Intern Med* 22:67–72.
- [24] Guidry, V. T., Rhodes, S. M., Woods, C. G., Hall, D. J., & Rinsky, J. L. (2018). Connecting environmental justice and community health: Effects of hog production in North Carolina. *North Carolina medical journal*, 79(5), 324-328.
- [25] Guillien, A., Puyraveau, M., Soumagne, T., et al., 2016. Prevalence and risk factors for COPD in farmers: a cross-sectional controlled study. *Eur. Respir. J.* 47, 95–103. doi: 10.1183/13993003.00153-2015.
- [26] Guillien, A., Soumagne, T., Dalphin, J.C., Degano, B., 2019. COPD, airflow limitation and chronic bronchitis in farmers: a systematic review and meta-analysis. *Occup. Environ. Med.* 76, 58–68. doi: 10.1136/oemed-2018-105310.
- [27] Hartung J., and Schulz J. 2008. Occupational and environmental risks caused by bioaerosols hazards of workers in swine confinement buildings. *J. Occup. Med.* 19: 383–387.
- [28] Hawley, B., Schaeffer, J., Poole, J. A., Dooley, G. P., Reynolds, S., & Volckens, J. (2015). Differential response of human nasal and bronchial epithelial cells upon exposure to size-fractionated dairy dust. *Journal of Toxicology and Environmental Health, Part A*, 78(9), 583-594.
- [29] Hisauchi-Kojima K, Sumi Y, Miyashita Y, Miyake S, Toyoda H, Kurup VP, et al. Purification of the antigenic components of pigeon dropping extract, the responsible agent for cellular immunity in pigeon breeder's disease. *J Allergy Clin Immunol* 1999;103: 1158–1165.
- [30] Hoppin, J. A., Umbach, D. M., London, S. J., Alavanja, M. C. R., & Sandler, D. P. (2003). Animal production and wheeze in the Agricultural Health Study: interactions with atopy, asthma, and smoking. *Occupational and environmental medicine*, 60(8), e3-e3.
- [31] Huy, T., K. deSchipper, M. Chan-Yeung, and S. Kennedy. 1991. Grain dust and lung function. *Am. Rev. Respir. Dis.* 144:1314–1321.
- [32] Iowa Concentrated Animal Feeding Operations Air Quality Study. Final Report. [http://library.state.or.us/repository/2012/201204101013082/appendix_L.pdf]. Accessed 10 Jul 2018.
- [33] J., Kirychuck, S., Reed, S., Schaeffer, J. W., Schenker, M. B., Schlünssen, V. and Kang, J. Y., Nan, X., Jin, M. S., Youn, S. J., Ryu, Y. H., Mah, S., ... & Lee, J. O. (2009). Recognition of lipopeptide patterns by Toll-like receptor 2-Toll-like receptor 6 heterodimer. *Immunity*, 31(6), 873-884.
- [34] Kang, J. Y., Nan, X., Jin, M. S., Youn, S. J., Ryu, Y. H., Mah, S., ... & Lee, J. O. (2009). Recognition of lipopeptide patterns by Toll-like receptor 2-Toll-like receptor 6 heterodimer. *Immunity*, 31(6), 873-884.
- [35] Kaur, G., & Sethi, R. S. (2020). Multiple exposures to poultry barn air and lipopolysaccharide synergistically increase the pulmonary expression of TLR-4 and IL-1β. *Journal of occupational health*, 62(1), e12094.
- [36] Kemper, N. (2008). Veterinary antibiotics in the aquatic and terrestrial environment. *Ecological indicators*, 8(1), 1-13.
- [37] Kim, D., Sass-Kortsak, A., Purdham, J. T., Dales, R. E., & Brook, J. R. (2006). Associations between personal exposures and fixed-site ambient measurements of fine particulate matter, nitrogen dioxide, and carbon monoxide in Toronto, Canada. *Journal of exposure science & environmental epidemiology*, 16(2), 172-183.
- [38] Kirkhorn, S. R., & Garry, V. F. (2000). Agricultural lung diseases. *Environmental health perspectives*, 108(suppl 4), 705-712.
- [39] Kirychuk, S. P., Reynolds, S. J., Koehncke, N. K., Lawson, J., Willson, P., Senthilselvan, A., ... & Dosman, J. A. (2010). Endotoxin and dust at respirable and nonrespirable particle sizes are not consistent between cage-and floor-housed poultry operations. *Annals of occupational hygiene*, 54(7), 824-832.
- [40] Kouimintzis, D., Chatzis, C. & Linos, A. Health effects of livestock farming in Europe. *J Public Health* 15, 245 (2007).
- [41] Kullman, G. J., Thorne, P. S., Waldron, P. F., Marx, J. J., Ault, B., Lewis, D. M., ... & Merchant, J. A. (1998). Organic dust exposures from work in dairy barns. *American Industrial Hygiene Association Journal*, 59(6), 403-413.
- [42] Lacey, J. (1994). Microorganisms in organic dust. *Organic dusts-Exposure, effects, and prevention. (Edited by R. Rylander and RR Jacobs)*, Lewis Publishers, London, 17-41.

- [43] Lee, W. Y. (2011). Clinical year-in-review of occupational lung disease. *Tuberculosis and Respiratory Diseases*, 71(5), 317-321.
- [44] Ley B, Newton CA, Arnould I, Elicker BM, Henry TS, Vittinghoff E, et al. The MUC5B promoter polymorphism and telomere length in patients with chronic hypersensitivity pneumonitis: an observational cohort-control study. *Lancet Respir Med* 2017;5:639–647.
- [45] Linaker, C., & Smedley, J. (2002). Respiratory illness in agricultural workers. *Occupational medicine*, 52(8), 451-459.
- [46] Lopez-Castejon, G., & Brough, D. (2011). Understanding the mechanism of IL-1 β secretion. *Cytokine & growth factor reviews*, 22(4), 189-195.
- [47] Lundin, J. I., & Checkoway, H. (2009). Endotoxin and cancer. *Environmental health perspectives*, 117(9), 1344-1350.
- [48] Marescaux, A., Degano, B., Soumagne, T., et al., 2016. Impact of farm modernity on the prevalence of chronic obstructive pulmonary disease in dairy farmers. *Occup. Environ. Med.* 73, 127–133. doi: 10.1136/oemed-2014-102697.
- [49] Martin, W. T., Zhang, Y., Willson, P., Archer, T. P., Kinahan, C., & Barber, E. M. (1996). Bacterial and fungal flora of dust deposits in a pig building. *Occupational and Environmental Medicine*, 53(7), 484-487.
- [50] Merchant, J., Miller, E., Campbell, J., Twigg, J., Marx, J., Ault, B., & Burmeister, L. (1991). Case-control assessment of lung function among dairy farmers. *Am. Rev. Resp. Dis*, 143.
- [51] Monsó, E., Riu, E., Radon, K., Magarolas, R., Danuser, B., Iversen, M., ... & Nowak, D. (2004). Chronic obstructive pulmonary disease in never-smoking animal farmers working inside confinement buildings. *American journal of industrial medicine*, 46(4), 357-362.
- [52] Nath Neerukonda, S., Mahadev-Bhat, S., Aylward, B., Johnson, C., Charavaryamath, C., & Arsenault, R. J. (2018). Kinome analyses of inflammatory responses to swine barn dust extract in human bronchial epithelial and monocyte cell lines. *Innate immunity*, 24(6), 366-381.
- [53] Nordgren, T. M., & Charavaryamath, C. (2018). Agriculture occupational exposures and factors affecting health effects. *Current allergy and asthma reports*, 18(12), 1-8.
- [54] Nordgren, T. M., Bauer, C. D., Heires, A. J., Poole, J. A., Wyatt, T. A., West, W. W., & Romberger, D. J. (2015). Maresin-1 reduces airway inflammation associated with acute and repetitive exposures to organic dust. *Translational Research*, 166(1), 57-69.
- [55] Nordgren, T. M., Heires, A. J., Wyatt, T. A., Poole, J. A., LeVan, T. D., Cerutis, D. R., & Romberger, D. J. (2013). Maresin-1 reduces the pro-inflammatory response of bronchial epithelial cells to organic dust. *Respiratory research*, 14(1), 1-10.
- [56] Omland, O. (2002). Exposure and respiratory health in farming in temperate zones-a review of the literature. *Annals of Agricultural and Environmental Medicine*, 9(2), 119-136.
- [57] Oosting, M., Cheng, S. C., Bolscher, J. M., Vesterling-Stenger, R., Plantinga, T. S., Verschuere, I. C., ... & Joosten, L. A. (2014). Human TLR10 is an anti-inflammatory pattern-recognition receptor. *Proceedings of the National Academy of Sciences*, 111(42), E4478-E4484.
- [58] Pardo A, Barrios R, Gaxiola M, Segura-Valdez L, Carrillo G, Estrada A, et al. Increase of lung neutrophils in hypersensitivity pneumonitis is associated with lung fibrosis. *Am J Respir Crit Care Med* 2000;161:1698–1704.
- [59] PastvaAM, WrightJR, Williams KL (2007) Immunomodulatory roles of surfactant proteins A and D: implications in lung disease. *Proc AmThorac Soc* 4:252–257.
- [60] Peters, S., Kromhout, H., Olsson, A. C., Wichmann, H. E., Brüske, I., Consonni, D., ... & Vermeulen, R. (2012). Occupational exposure to organic dust increases lung cancer risk in the general population. *Thorax*, 67(2), 111-116.
- [61] Pfister, H., Madec, L., Cann, P.L., et al., 2018. Factors determining the exposure of dairy farmers to thoracic organic dust. *Environ. Res.* 165, 286–293. doi: 10.1016/j.envres.2018.04.031.
- [62] Poole, J. A., Alexis, N. E., Parks, C., MacInnes, A. K., Gentry-Nielsen, M. J., Fey, P. D., ... & Romberger, D. J. (2008). Repetitive organic dust exposure in vitro impairs macrophage differentiation and function. *Journal of allergy and clinical immunology*, 122(2), 375-382.
- [63] Poole, J. A., Wyatt, T. A., Kielian, T., Oldenburg, P., Gleason, A. M., Bauer, A., ... & Romberger, D. J. (2011). Toll-like receptor 2 regulates organic dust-induced airway inflammation. *American journal of respiratory cell and molecular biology*, 45(4), 711-719.
- [64] Poole, J.A., Romberger, D.J., 2012. Immunological and inflammatory responses to organic dust in agriculture. *Curr. Opin. Allergy Clin. Immunol.* 12, 126–132. doi: 10.1097/ACI.0b013e3283511d0e.
- [65] Preller, L., Heederik, D., Kromhout, H., Boleij, J. S., & Tielen, M. J. (1995). Determinants of dust and endotoxin exposure of pig farmers: development of a control strategy using empirical modelling. *The Annals of occupational hygiene*, 39(5), 545-557.
- [66] Reynolds, S. J., Nonnenmann, M. W., Basinas, I., Davidson, M., Elfman, L., Gordon, J., ... & Sigsgaard, T. (2013). Systematic review of respiratory health among dairy workers. *Journal of agromedicine*, 18(3), 219-243.
- [67] Romberger, D. J., Heires, A. J., Nordgren, T. M., Souder, C. P., West, W., Liu, X. D., ... & Wyatt, T. A. (2015). Proteases in agricultural dust induce lung inflammation through PAR-1 and PAR-2 activation. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 309(4), L388-L399.
- [68] Rylander, R., & Jacobs, R. R. (1994). *Organic dusts exposure, effects, and prevention*. CRC Press.
- [69] Sahlander, K., Larsson, K., & Palmberg, L. (2010). Altered innate immune response in farmers and smokers. *Innate immunity*, 16(1), 27-38.
- [70] Schenker, M. (2000). Exposures and health effects from inorganic agricultural dusts. *Environmental health perspectives*, 108(suppl 4), 661-664.
- [71] Schenker, M. B., Christiani, D., Cormier, Y., Dimich-Ward, H., Doekes, G., Dosman, J., ... & Chan-Yeung, M. (1998).

- Respiratory health hazards in agriculture. *American journal of respiratory and critical care medicine*, 158(5), S1-S76.
- [72] Schneberger, D., Aulakh, G., Channabasappa, S., & Singh, B. (2016). Toll-like receptor 9 partially regulates lung inflammation induced following exposure to chicken barn air. *Journal of Occupational Medicine and Toxicology*, 11(1), 1-10.
- [73] Schneberger, D., DeVasure, J. M., Kirychuk, S. A., & Wyatt, T. A. (2018). Organic barn dust inhibits surfactant protein D production through protein kinase-c alpha dependent increase of GPR116. *PloS one*, 13(12), e0208597.
- [74] Senthilselvan, A., Dosman, J. A., Chénard, L., Burch, L. H., Predicala, B. Z., Sorowski, R., ... & Schwartz, D. A. (2009). Toll-like receptor 4 variants reduce airway response in human subjects at high endotoxin levels in a swine facility. *Journal of allergy and clinical immunology*, 123(5), 1034-1040.
- [75] Senthilselvan, A., Dosman, J. A., Kirychuk, S. P., Barber, E. M., Rhodes, C. S., Zhang, Y., & Hurst, T. S. (1997). Accelerated lung function decline in swine confinement workers. *Chest*, 111(6), 1733-1741.
- [76] Sethi, R. S., Schneberger, D., & Singh, B. (2013). Characterization of the lung epithelium of wild-type and TLR9^{-/-} mice after single and repeated exposures to chicken barn air. *Experimental and Toxicologic Pathology*, 65(4), 357-364.
- [77] Sethi, R. S., Schneberger, D., Charavaryamath, C., & Singh, B. (2017). Pulmonary innate inflammatory responses to agricultural occupational contaminants. *Cell and tissue research*, 367(3), 627-642.
- [78] Sigsgaard, T., Basinas, I., Doekes, G., de Blay, F., Folletti, I., Heederik, D., ... & Siracusa, A. (2020). Respiratory diseases and allergy in farmers working with livestock: a EAACI position paper. *Clinical and Translational Allergy*, 10(1), 1-30.
- [79] Sigsgaard, T., Omland, Ø., & Thorne, P. S. (2010). Asthma-like diseases in agriculture. In *Occupational Asthma* (pp. 163-183). Birkhäuser Basel.
- [80] Skerrett SJ, Liggitt HD, Hajjar AM, Ernst RK, Miller SI, Wilson CB. Respiratory epithelial cells regulate lung inflammation in response to inhaled endotoxin. *Am J Physiol Lung Cell Mol Physiol*. 2004;287(1): L143-L152.
- [81] Smid, T., Heederik, D., Houba, R., & Quanjer, P. H. (1992). Dust-and endotoxin-related respiratory effects in the animal feed industry. *American journal of respiratory and critical care medicine*, 146(6), 1474-1479.
- [82] Soumagne, T., Degano, B., Guillien, A., Annesi-Maesano, I., Andujar, P., Hue, S., ... & Dalphin, J. C. (2020). Characterization of chronic obstructive pulmonary disease in dairy farmers. *Environmental Research*, 188, 109847.
- [83] Strieter, R. M., Belperio, J. A., & Keane, M. P. (2002). Cytokines in innate host defense in the lung. *The Journal of clinical investigation*, 109(6), 699-705.
- [84] Sugiyama, K. I., Muroi, M., & Tanamoto, K. I. (2008). A novel TLR4-binding peptide that inhibits LPS-induced activation of NF-κB and in vivo toxicity. *European journal of pharmacology*, 594(1-3), 152-156.
- [85] Szczyrek, M., Krawczyk, P., Milanowski, J., Jastrzebska, I., Zwolak, A., & Daniluk, J. (2011). Chronic obstructive pulmonary disease in farmers and agricultural workers-an overview. *Annals of Agricultural and Environmental Medicine*, 18(2).
- [86] Takeda, K., Kaisho, T., & Akira, S. (2003). Toll-like receptors. *Annual review of immunology*, 21(1), 335-376.
- [87] Taluja, M. K., Gupta, V., Sharma, G., & Arora, J. S. (2019). Respiratory Hazards to Occupational Exposure of Poultry Dust in Poultry Farm Workers in Northern India. *Indian J PhysiolPharmacol*, 63(3), 223-230.
- [88] Tsan, M. F., & Gao, B. (2004). Endogenous ligands of Toll-like receptors. *Journal of leukocyte biology*, 76(3), 514-519.
- [89] Van Dijk, C. E., Garcia-Aymerich, J., Carsin, A. E., Smit, L. A., Borlée, F., Heederik, D. J., ... & Zock, J. P. (2016). Risk of exacerbations in COPD and asthma patients living in the neighbourhood of livestock farms: observational study using longitudinal data. *International journal of hygiene and environmental health*, 219(3), 278-287.
- [90] Vasakova M, Selman M, Morell F, Sterclova M, Molina-Molina M, Raghu G. Hypersensitivity pneumonitis: current concepts of pathogenesis and potential targets for treatment. *Am J Respir Crit Care Med* 2019;200:301–308.
- [91] Vernooij, J. H., Dentener, M. A., Van Suylen, R. J., Buurman, W. A., & Wouters, E. F. (2002). Long-term intratracheal lipopolysaccharide exposure in mice results in chronic lung inflammation and persistent pathology. *American journal of respiratory cell and molecular biology*, 26(1), 152-159.
- [92] Viegas, S., Caetano, L. A., Korkalainen, M., Faria, T., Pacifico, C., Carolino, E., ... & Viegas, C. (2017). Cytotoxic and inflammatory potential of air samples from occupational settings with exposure to organic dust. *Toxics*, 5(1), 8.
- [93] Viegas, S., Mateus, V., Almeida-Silva, M., Carolino, E., & Viegas, C. (2013). Occupational exposure to particulate matter and respiratory symptoms in Portuguese swine barn workers. *Journal of Toxicology and Environmental Health, Part A*, 76(17), 1007-1014.
- [94] Woldeamanuel, G. G., Mingude, A. B., Yitbarek, G. Y., & Taderegew, M. M. (2020).
- [95] Zähringer, U., Lindner, B., Inamura, S., Heine, H., & Alexander, C. (2008). TLR2—promiscuous or specific? A critical re-evaluation of a receptor expressing apparent broad specificity. *Immunobiology*, 213(3-4), 205-224.
- [96] Zejda, J. E., McDuffie, H. H., & Dosman, J. A. (1993). Epidemiology of health and safety risks in agriculture and related industries. Practical applications for rural physicians. *Western journal of medicine*, 158(1), 56.